



COLO-205

CSI314Hu11

Instruction manual

FOR RESEARCH USE ONLY
NOT FOR USE IN CLINICAL DIAGNOSTIC PROCEDURES

2nd Edition (Revised in Apr, 2025)

[DESCRIPTION]

The COLO 205 cell line is made up of epithelial cells isolated in 1975 from ascitic fluid derived from a 70-year-old, White, male with colon cancer. The cells can be used for cancer and toxicology research.

Synonyms: Colo 205; CoLo 205; COLO-205; Colo-205; COLO.205; Colo205; COLO205; Co 205

Organism: Homo sapiens, human

Tissue Source: Large intestine; Colon, ascites metastasis

Disease: Adenocarcinoma; Colorectal; Dukes' type D

Gender: Male

Age: 70 years

Cell Type: Epithelial

Growth Properties: Mixed, adherent and suspension

[PROPERTIES]

Cell activity: >85% (Viability by Trypan Blue Exclusion).

Formulation: Frozen 1 mL or T25 flask.

Biosafety: Negative for HIV-1, HBV, HCV, mycoplasma, bacteria, yeast and fungi.

Applications: For research use only. It is not approved for human or animal use, or for application in clinical diagnostic procedures.

Size: >5×10⁵cell/vial

[STORAGE]

Upon receiving, check all containers for leakage or breakage. directly and immediately transfer the cells from dry ice to liquid nitrogen and keep the cells in liquid nitrogen until they are needed for experiments.

Form & Buffer: Supplied as solution form in frozen stock solution, containing 50% base medium +40%FBS+10%DMSO.

Storage conditions: liquid nitrogen

[USAGE]

Culture conditions:

Complete growth medium: RPMI-1640+10%FBS+1%Penicillin-Streptomycin Solution

Temperature: 37°C

Condition: 95% air, 5% carbon dioxide



Cell recovery:

1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the cap out of the water. The thawing time is about 2 minutes.
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by spraying with 75% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
3. Transfer the vial contents to a centrifuge tube containing 9.0mL complete culture medium. and spin at approximately 1000 rpm for 5 minutes.
4. Resuspend cell pellet with the recommended complete medium . and dispense into a T25 culture flask.
5. Incubate the culture at 37°C, 5% CO₂ in a suitable incubator.

Cell passage:

Shake the flask to transfer the suspended cells into the centrifuge tube, retaining the suspended cells. The cells that are still attached can be removed using trypsin digestion and combined with the remaining floating cells.

1. Briefly rinse the cell layer with 0.25% (w/v) Trypsin-0.53 mM EDTA solution to remove all traces of serum which, contains trypsin inhibitor.
2. Add 1.0 mL of Trypsin-EDTA solution to flask and observe cells under an inverted microscope until cell layer is dispersed (usually within 2 to 3 minutes. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal). Stop digestion by adding 2-3 ml of complete medium containing 10% serum. Make it a single cell suspension.
3. Add appropriate the cell suspension to the centrifuge tube containing the floating cells. Centrifuge at 1000 rpm for 5 minutes.
4. Add the fresh medium to resuspend the cells. Unless otherwise stated, the recommended ratio of subcultivation is 1/2-1/5.

[Shipping]

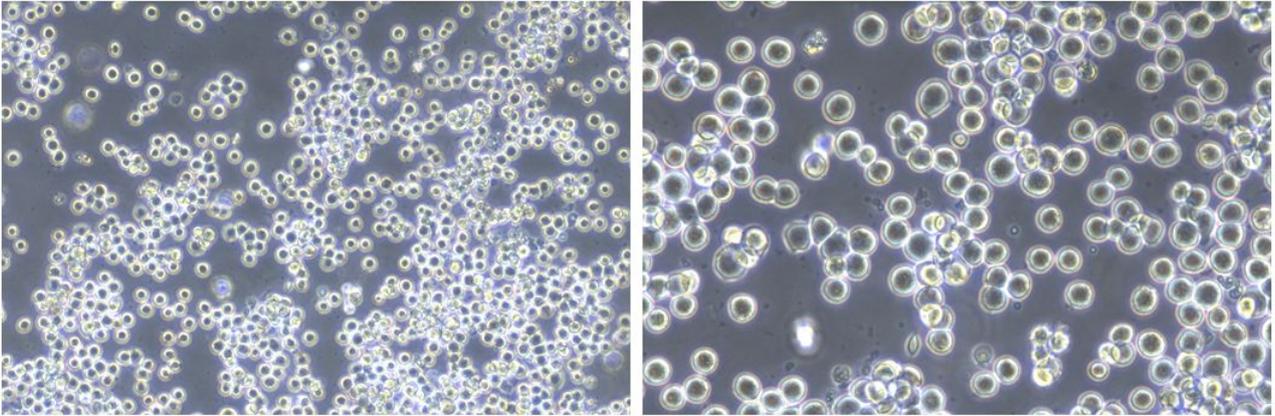
Dry ice.

[IMPORTANTNOTE]

1. This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.
2. To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.
3. After cell recovery, please take regular microscopic examination and photos to record the growth status of cells.
4. Read the instructions carefully, and keep and operate in strict accordance with the instructions. If you observe abnormalities or have questions about cell culture operations, please contact us in time.



[Figure]



Morphology of COLO-205 (Optical microscope, 100x, 200x)