



Jurkat, Clone E6-1

CSI185Hu11

Instruction manual

FOR RESEARCH USE ONLY
NOT FOR USE IN CLINICAL DIAGNOSTIC PROCEDURES

1st Edition (Revised in Oct, 2023)

[DESCRIPTION]

Jurkat, Clone E6-1 is a clone of the Jurkat-FHCRC cell line, a derivative of the Jurkat cell line, which was established from the peripheral blood of a 14-year-old, male, acute T-cell leukemia patient. This cell line can be used in immune system disorder research and immunology and immuno-oncology research.

Synonyms: Jurkat E6-1; Jurkat Clone E6-1; Jurkat (clone E6-1)

Organism: Homo sapiens, human

Tissue Source: Peripheral blood

Disease: Acute T cell leukemia

Cell Type: T lymphoblast

Morphology: Lymphoblast

Growth Properties: Suspension

[PROPERTIES]

Cell activity: >95% (Viability by Trypan Blue Exclusion).

Formulation: Frozen 1 mL or T25 flask.

Biosafety: Negative for HIV-1, HBV, HCV, mycoplasma, bacteria, yeast and fungi.

Applications: For research use only. It is not approved for human or animal use, or for application in clinical diagnostic procedures.

Size: >5×10⁵cell/vial

[STORAGE]

Upon receiving, check all containers for leakage or breakage. directly and immediately transfer the cells from dry ice to liquid nitrogen and keep the cells in liquid nitrogen until they are needed for experiments.

Form & Buffer: Supplied as solution form in frozen stock solution, containing 50% base medium +40%FBS+10%DMSO.

Storage conditions: liquid nitrogen

[USAGE]

Culture conditions:

Complete growth medium: RPMI-1640+10%FBS+1%Penicillin-Streptomycin Solution

Temperature: 37°C



Condition: 95% air, 5% carbon dioxide

Cell recovery:

1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the cap out of the water. The thawing time is about 2 minutes.
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by spraying with 75% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
3. Transfer the vial contents to a centrifuge tube containing 9.0mL complete culture medium. and spin at approximately 1200 rpm for 3 minutes.
4. Resuspend cell pellet with the recommended complete medium . and dispense into a T25 culture flask.
5. Incubate the culture at 37°C, 5% CO₂ in a suitable incubator.

Cell passage:

The cells are suspended cells, and maintain cultures at a cell concentraion between 1×10^5 and 1×10^6 viable cells/mL. Do not allow the cell density to exceed 3×10^5 cells/mL. Cultures can be maintained by the addition of fresh medium or replacement of medium. Alternatively, cultures can be established by centrifugation with subsequent resuspension at 1×10^5 viable cells/mL, and centrifugal speed reference 1200 rpm centrifuge 3 min. According to culture experience, it is recommended to use the "half-change solution method" for passage, that is, directly add an equal amount of fresh culture solution to the cell culture bottle, then the cells are blown evenly and transferred to two new T25 culture bottles for further culture. Depending on cell density, it is recommended to add fresh medium every 2-3 days.

[Shipping]

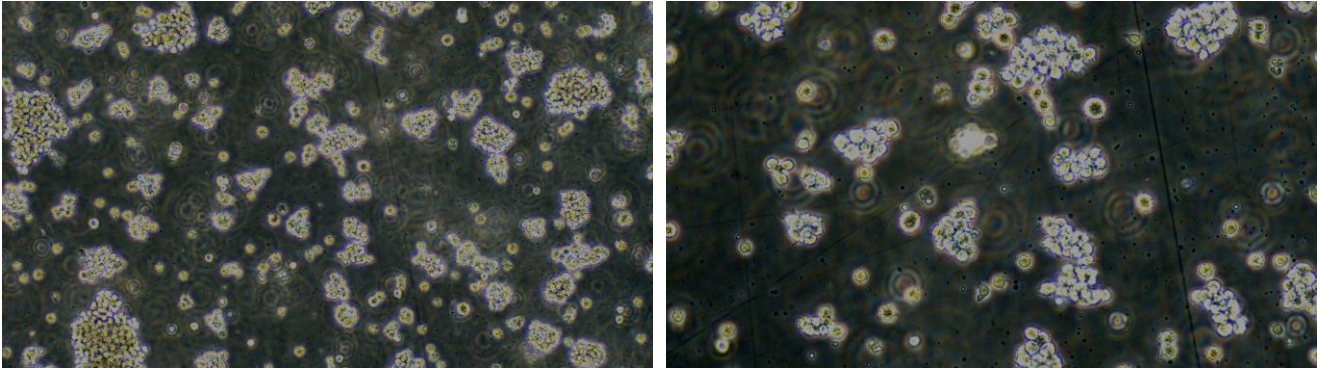
Dry ice.

[IMPORTANTNOTE]

1. This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.
2. To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.
3. After cell recovery, please take regular microscopic examination and photos to record the growth status of cells.
4. Read the instructions carefully, and keep and operate in strict accordance with the instructions. If you observe abnormalities or have questions about cell culture operations, please contact us in time.



[Figure]



Morphology of Jurkat, Clone E6-1 (Optical microscope, 100x, 200x)