



CSI083Rb01

Primary Rabbit Bone Marrow-derived Mesenchymal Stem Cells (BMSCs)

Organism Species: Oryctolagus cuniculus (Rabbit)

Instruction manual

FOR RESEARCH USE ONLY

NOT FOR USE IN CLINICAL DIAGNOSTIC PROCEDURES

2nd Edition (Revised in May, 2023)

[DESCRIPTION]

Cell Type: Mesenchymal Stem Cells

Synonyms: BMSCs

Species: Oryctolagus cuniculus (Rabbit)

Tissue Source: Bone marrow

Size: $>5 \times 10^5$ cell/mL

[PROPERTIES]

Cell activity: $>85\%$ (Viability by Trypan Blue Exclusion).

Formulation: Frozen 1 mL or T25 flask.

Biosafety: Negative for HIV-1, HBV, HCV, mycoplasma, bacteria, yeast and fungi.

Applications: For research use only. It is not approved for human or animal use, or for application in clinical diagnostic procedures.

Growth Properties: Adherent

[CONTENTS]

Form & Buffer: Supplied as solution form in frozen stock solution, containing 90% FBS+10% DMSO.

[USAGE]

Upon receiving the cells in a T-25 flask at room temperature, immediately transfer the cells to 37°C, 5% incubator; the cells in vials, directly and immediately transfer the cells from dry ice to liquid nitrogen.

Culture conditions:

DMEM/F12+10%FBS+1% Mesenchymal Stem Cell Growth Supplement+1%Penicillin-Streptomycin Solution

Temperature: 37°C

Condition: 95% air, 5% carbon dioxide

Cell recovery:

After receiving the cells, shake at 37°C in a water bath until completely dissolved, transfer to a 15 ml centrifuge tube, add 3-5 times complete culture solution, 1000 rpm for 5 min, discard the supernatant, and place in a T25 flask for culture.

Cell passage:



1. Cell passage when cell growth at 85-95%.
2. Discard the medium and wash with PBS 1-2 times.
3. Add 1 ml of Trypsin at 37°C, observe the cell under the microscope. If the cells are retracted and rounded, pat the culture flask to let the cells fall off. Stop digestion by adding 2 ml of complete medium containing 10% serum. Make it a single cell suspension.
4. Add the fresh medium to resuspend the cells. Unless otherwise stated, the recommended ratio of primary cells is 1/3.

[Shipping]

Dry ice.

[STORAGE]

Upon receiving, directly and immediately transfer the cells from dry ice to liquid nitrogen and keep the cells in liquid nitrogen until they are needed for experiments.

[IMPORTANT NOTE]

The cell is for research use only, we will not be responsible for any issue if the cell was used in clinical diagnostic or any other procedures.

[Figure]

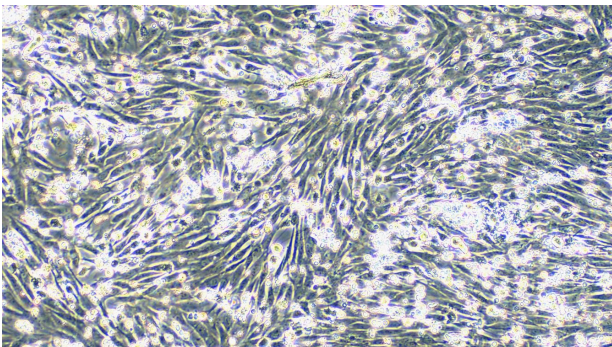


Figure 1

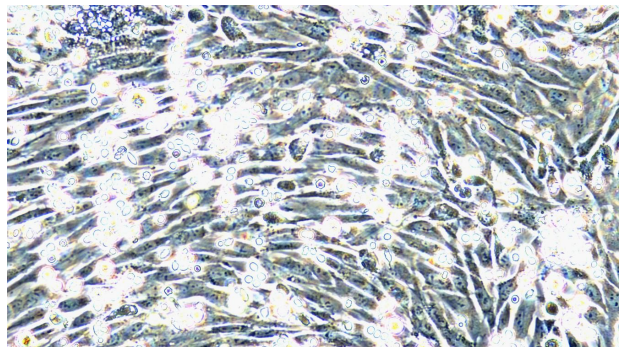


Figure 2

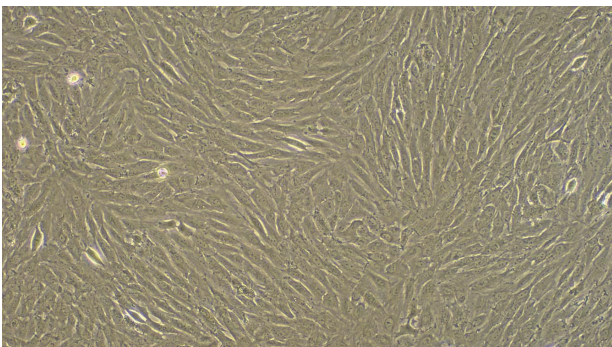


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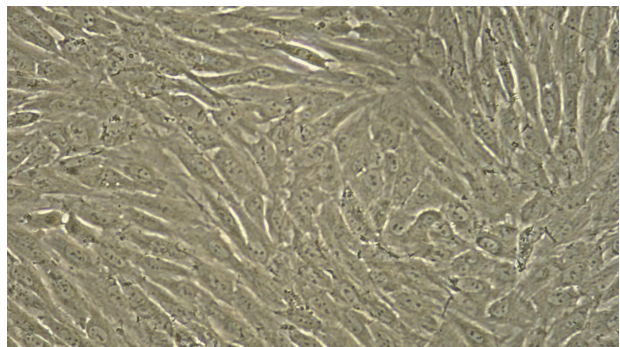


Figure 4

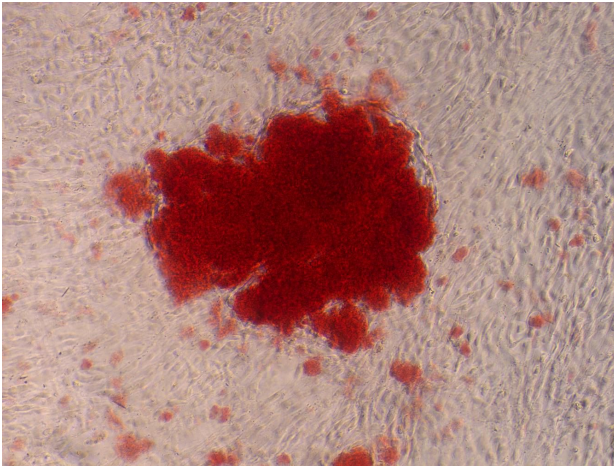


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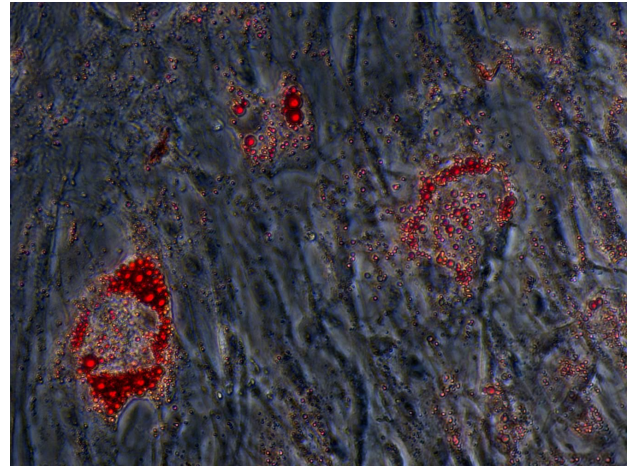


Figure 6

Figure 1 Morphology of BMMSCs on day 3 of primary culture (Optical microscope, ×100)

Figure 2 Morphology of BMMSCs on day 3 of primary culture (Optical microscope, ×200)

Figure 3 Morphology of P2 BMMSCs (Optical microscope, ×100)

Figure 4 Morphology of P2 BMMSCs (Optical microscope, ×200)

Figure 5 Alizarin red staining after osteogenic induction (Optical microscope, ×100)

Figure 6 Oil red O staining after adipogenic induction (Optical microscope, ×400)

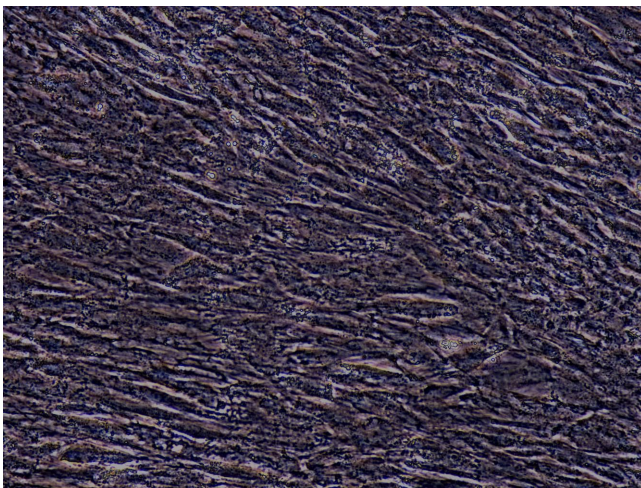


Figure 7

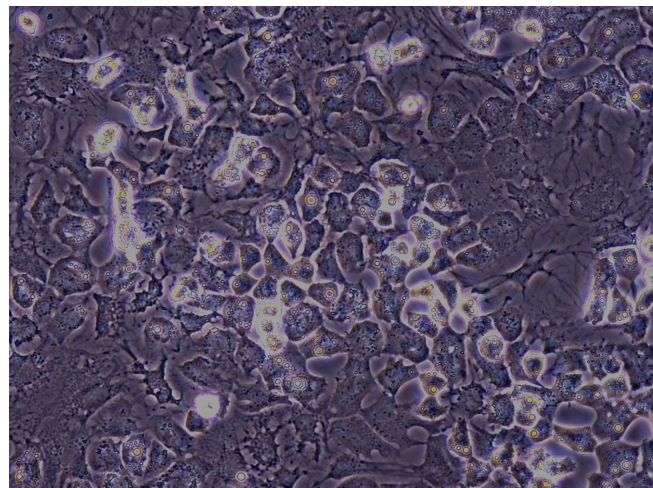


Figure 8

Figure 7 Control group: normal culture for 21 days (Optical microscope, ×200)

Figure 8 Induction group: chondroblast differentiation was induced for 21 days (Optical microscope, ×200)

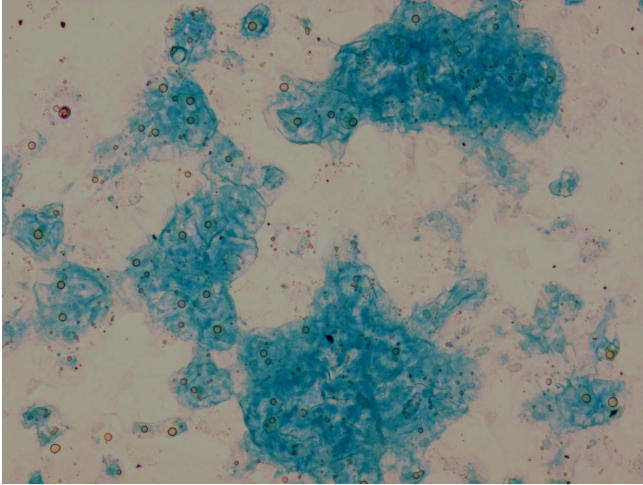


Figure 9

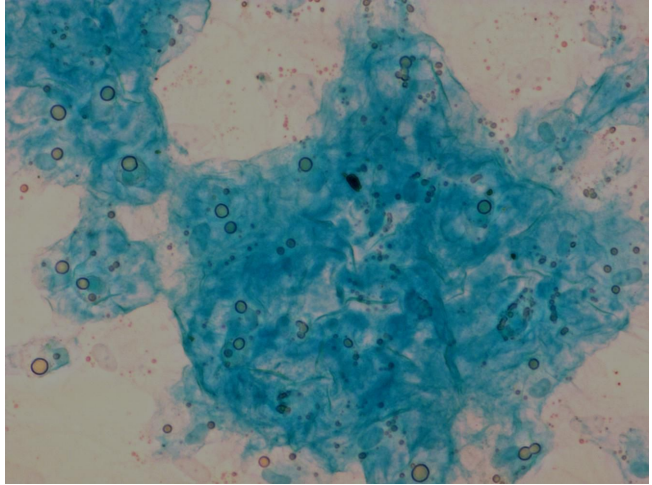


Figure 10

Figure 8 Alcian blue staining after chondroblast induction (Optical microscope,×200)

Figure 9 Alcian blue staining after chondroblast induction (Optical microscope,×400)

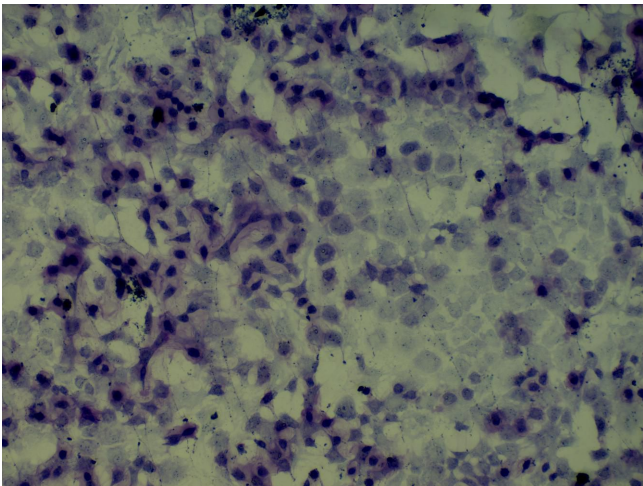


Figure 11

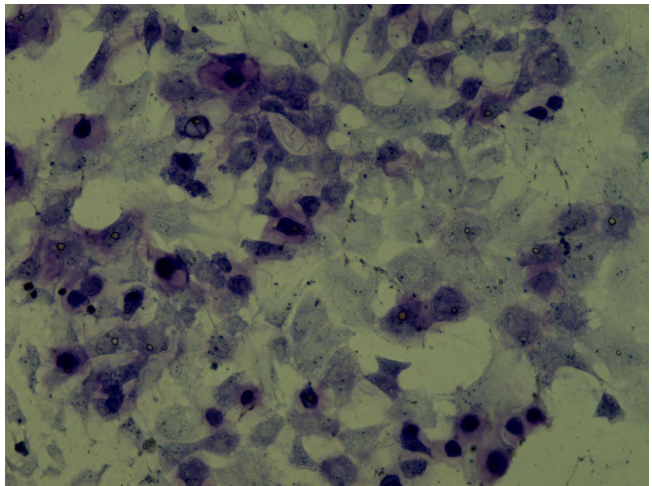


Figure 12

Figure 11 Toluidine blue staining after chondroblast induction (Optical microscope,×100)

Figure 12 Toluidine blue staining after chondroblast induction (Optical microscope,×200)